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## Resistenz gegen Schwermetalle und Interaktion mit Antibiotika-Resistenzen

# Christopher Rensing PhD

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University  
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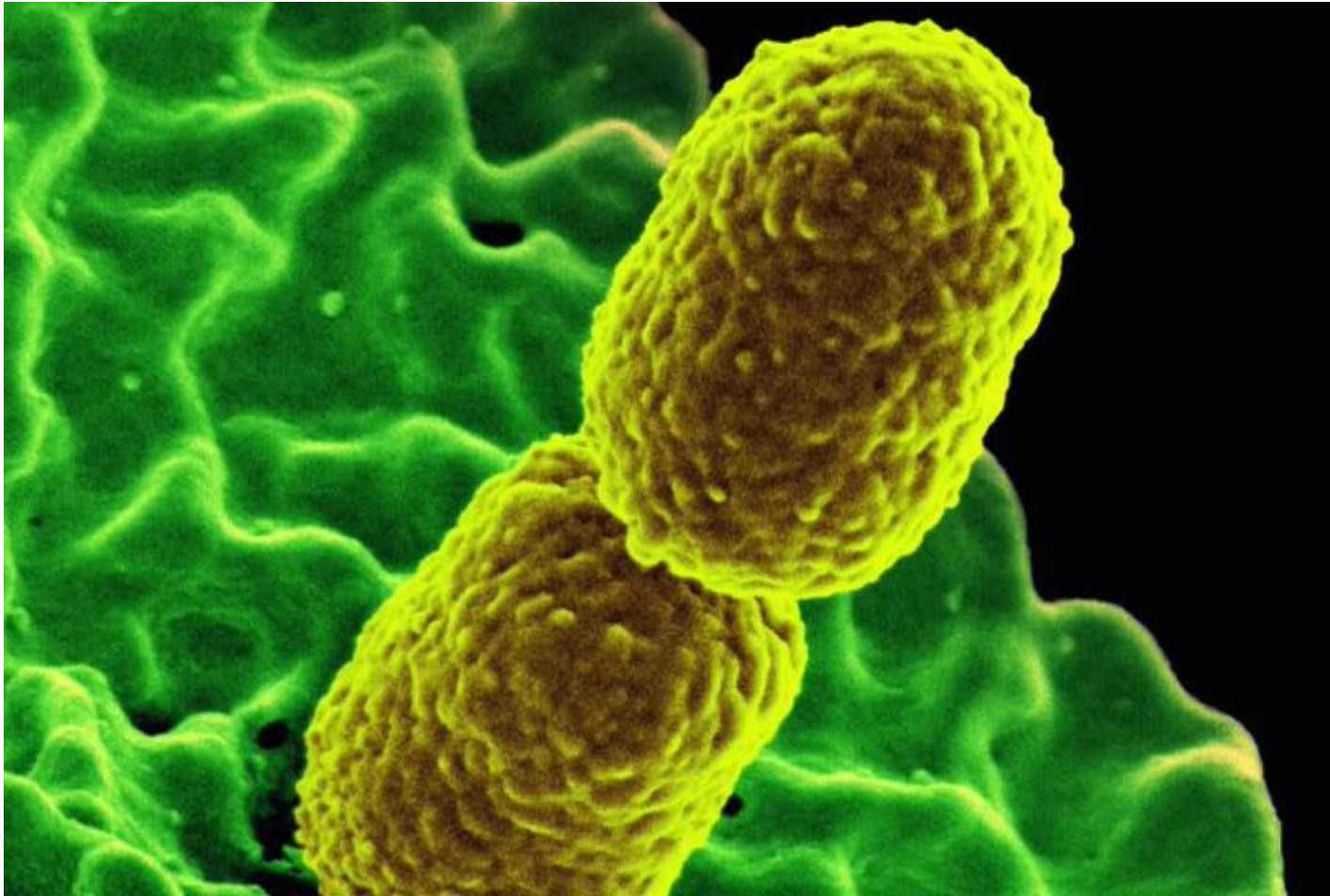


# Team





# *Klebsiella pneumoniae*



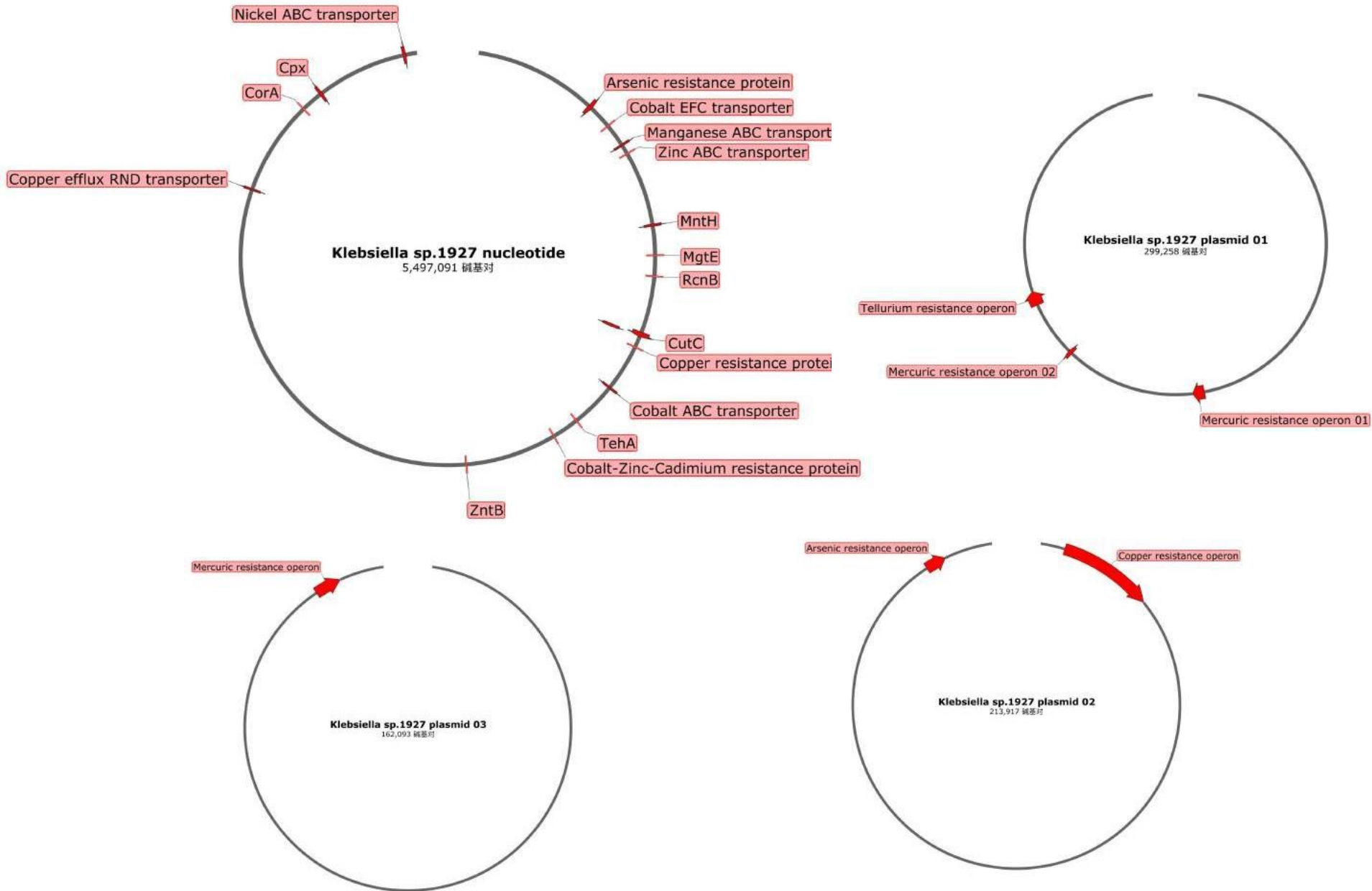
# The MIC of 1927 and 1954

	1927	1954
<b>Amp</b> (ppm)	>30,000	>30,000
<b>Km</b> (ppm)	13,000	14,000
<b>Cm</b> (ppm)	2,000	4,000
<b>Gm</b> (ppm)	6,000	7,000
<b>Tet</b> (ppm)	250	250
<b>Rif</b> (ppm)	10	15
<b>Str</b> (ppm)	40	10
<b>Au</b> ( $\mu$ mol/L)	900	<400
<b>Cu</b> (mmol/L)	2	0.25
<b>Ag</b> ( $\mu$ mol/L)	128	>1024
<b>Cd</b> (mmol/L)	7	5
<b>As (III)</b> (mmol/L)	12.5	0.5

1927 isolated from sputum of a patient with pneumonia.

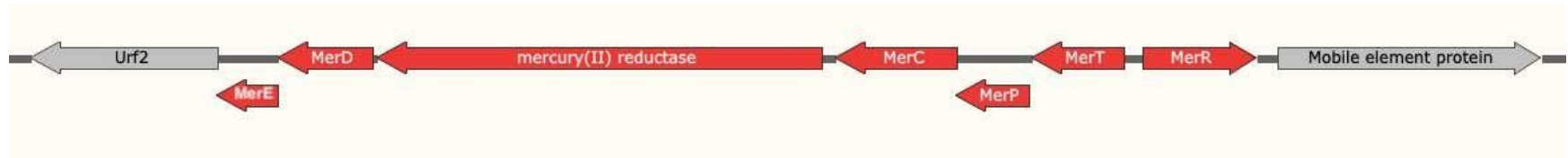
1954 isolated from sputum of a patient with hydrocephalus.

# *Klebsiella* sp.1927 Gene Map

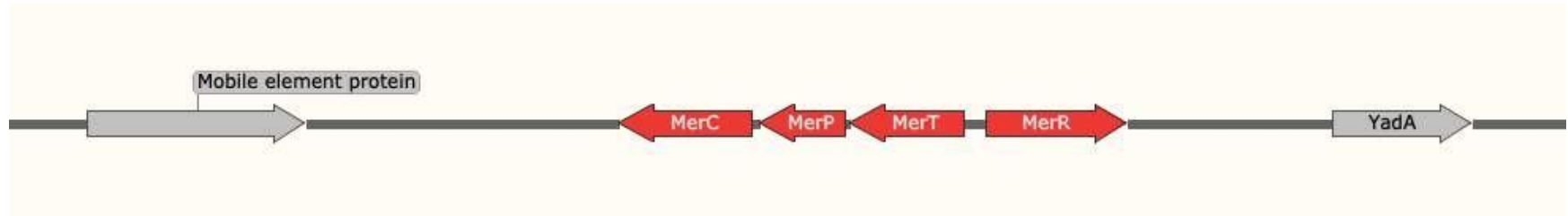


# Heavy metal resistance genes(cluster) in *Klebsiella* sp.1927 plasmid 01

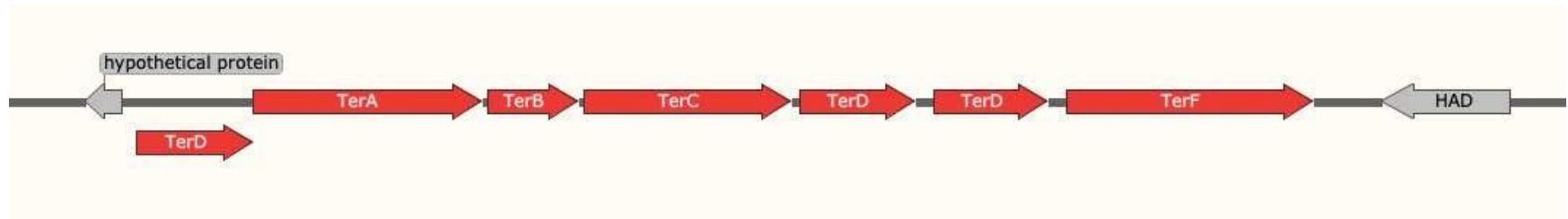
## Mercuric resistance operon 01



## Mercuric resistance operon 02

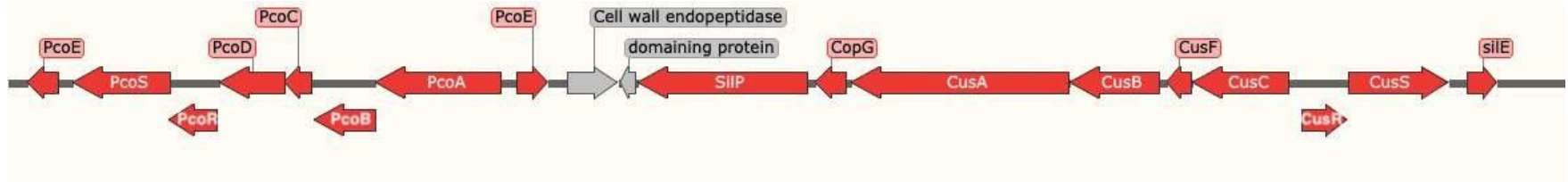


## Tellurium resistance operon

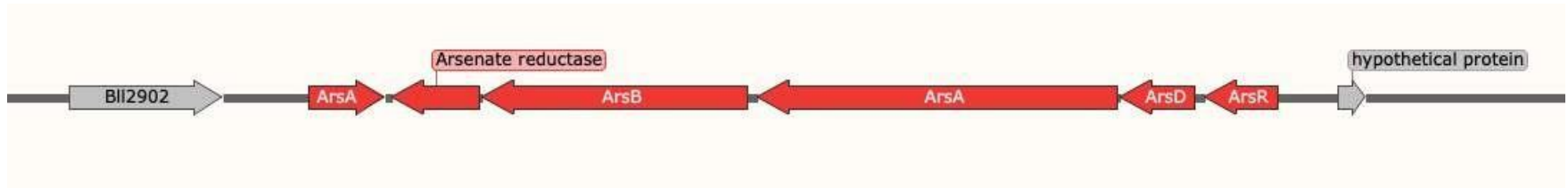


# Heavy metal resistance genes(cluster) in *Klebsiella* sp.1927 plasmid 02

## Copper resistance operon



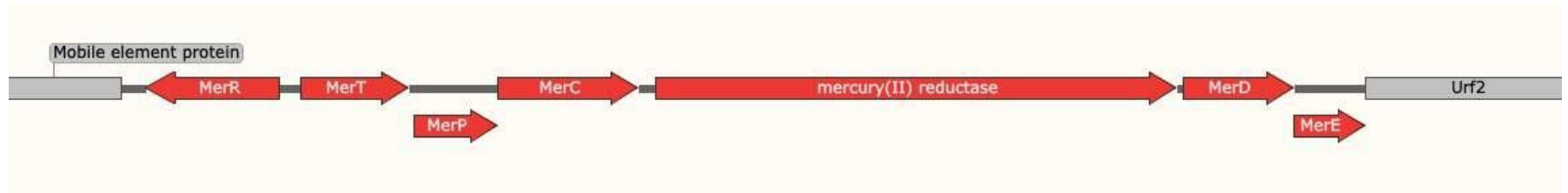
## Arsenic resistance operon





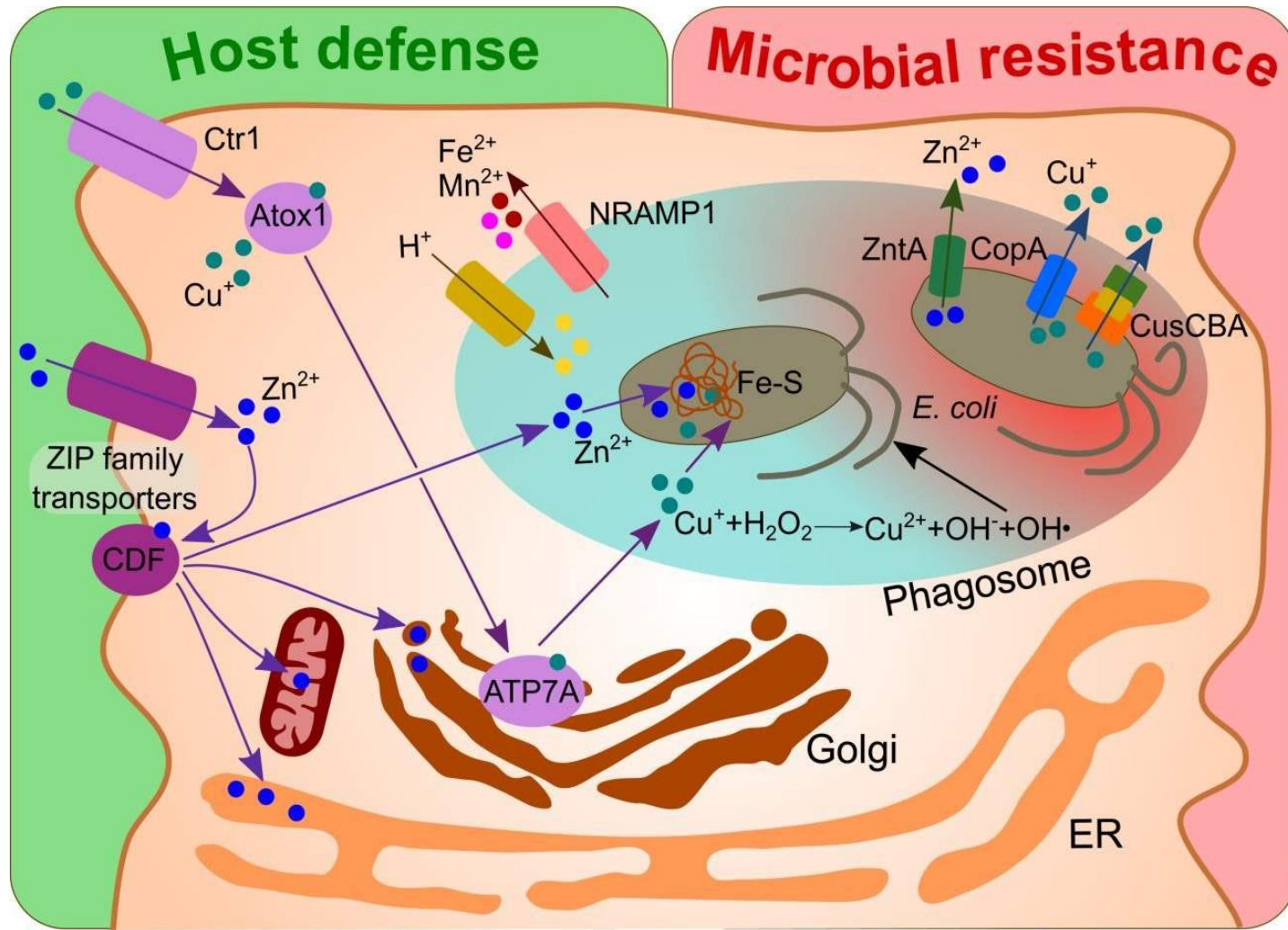
# Heavy metal resistance genes(cluster) in *Klebsiella* sp.1927 plasmid 03

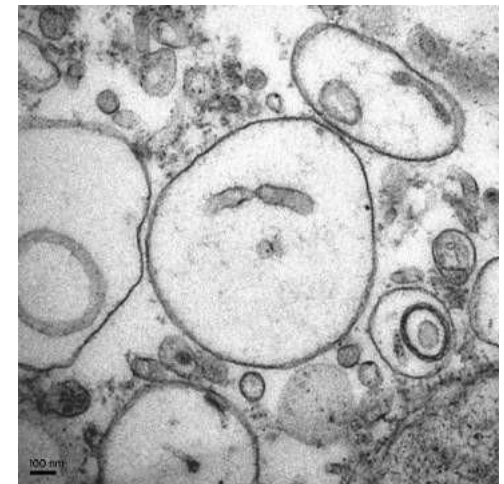
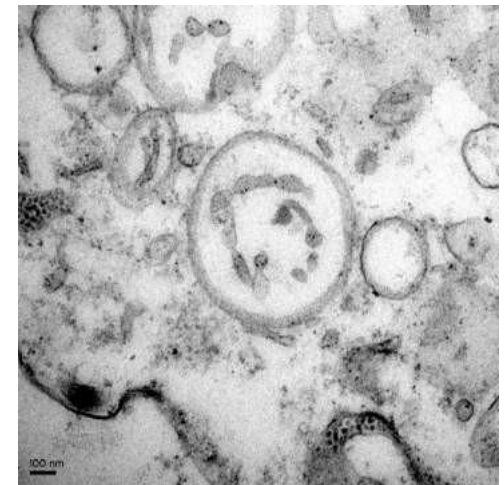
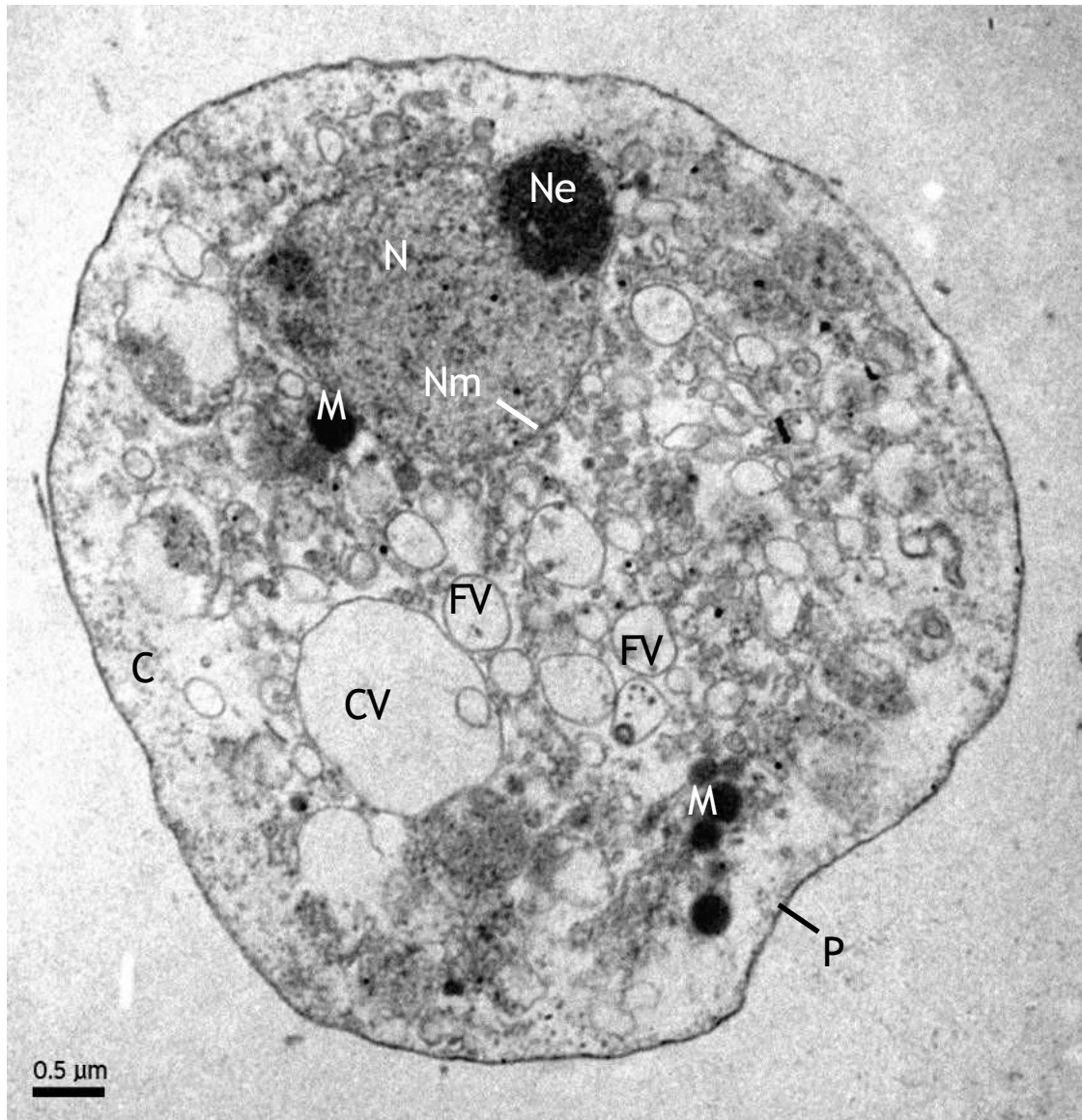
## Mercuric resistance operon



- **Warum?**
- **3 *mer* operons**
- **Tellurium operon**
- **19 Gen Kupfer Resistenz cluster**
- **Extra *ars* operon**

# Bacterial killing in macrophage/amoeba





**Ne:** nucleolus; **N:** nucleus; **Nm:** nucleus membrane; **M:** mitochondrion;  
**V:** Food vacuole; **CV:** contractile vacuole; **C:** Cytoplasm; **P:** Plasma membrane

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# Bacterial killing in macrophages and amoeba: do they all use a brass dagger?

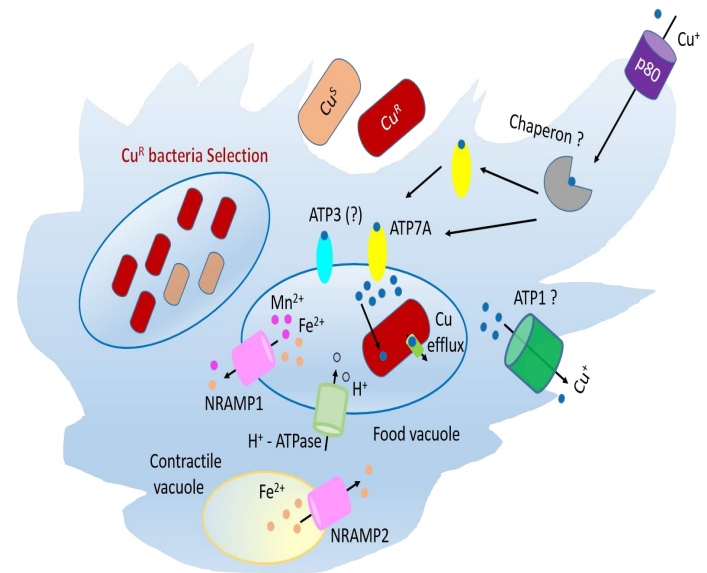
Nadezhda German<sup>1</sup>, Dominik Doyscher<sup>2</sup> & Christopher Rensing<sup>\*3</sup>

<sup>1</sup>Research Triangle Institute, Research Triangle Park, NC 27709, USA

<sup>2</sup>Department of Veterinary Sciences, Ludwig-Maximilians-University Munich, Oberschlesheim, Germany

<sup>3</sup>Department of Plant & Environmental Sciences, University of Copenhagen, Frederiksberg, Denmark

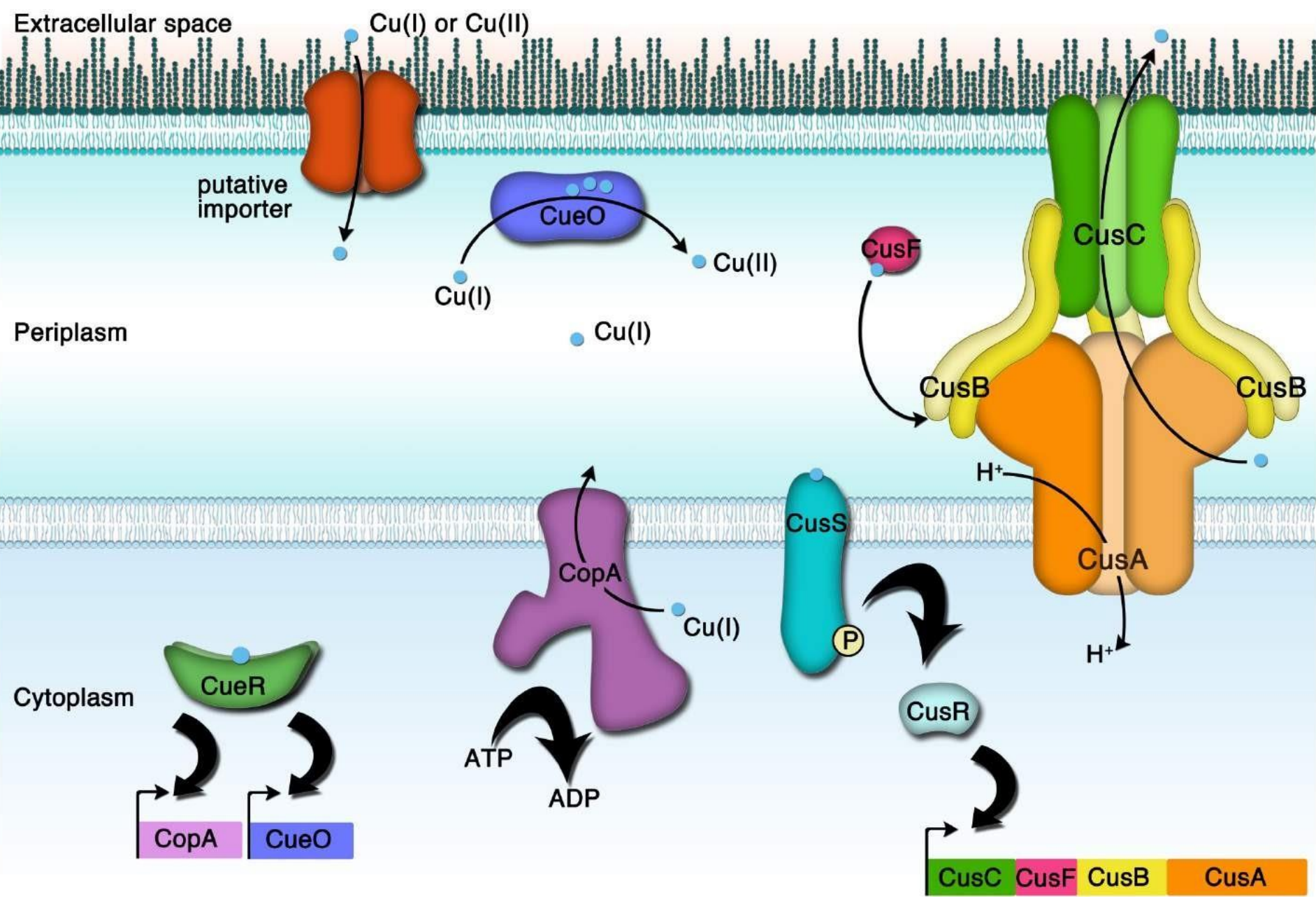
\*Author for correspondence: [chres@life.ku.dk](mailto:chres@life.ku.dk)



## Conclusions

This review presents a now considerable body of data consistent with the hypothesis that Cu and Zn toxicity can contribute to the clearance of bacterial pathogens. This has led to the intriguing concept of the “brass dagger” in innate immunity (30), which then raises the question of how such a bactericidal mechanism could be integrated into the complex immune sys-





Cytoplasm

Periplasm

Extracellular space

CopA CueO

CusC CusF CusB CusA

ATP ADP

H<sup>+</sup>

H<sup>+</sup>

Cu(I)

Cu(II)

Cu(I)

Cu(I)

Cu(I) or Cu(II)

putative importer

CueO

CopA

CusS

CusR

CusA

CusB

CusC

CusF

CusB

CusA

CusB

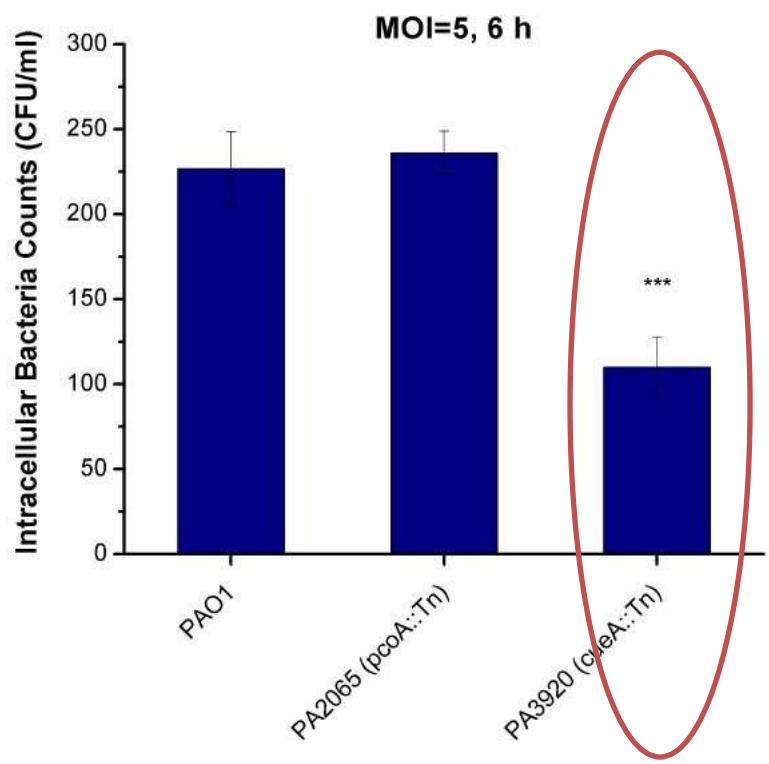
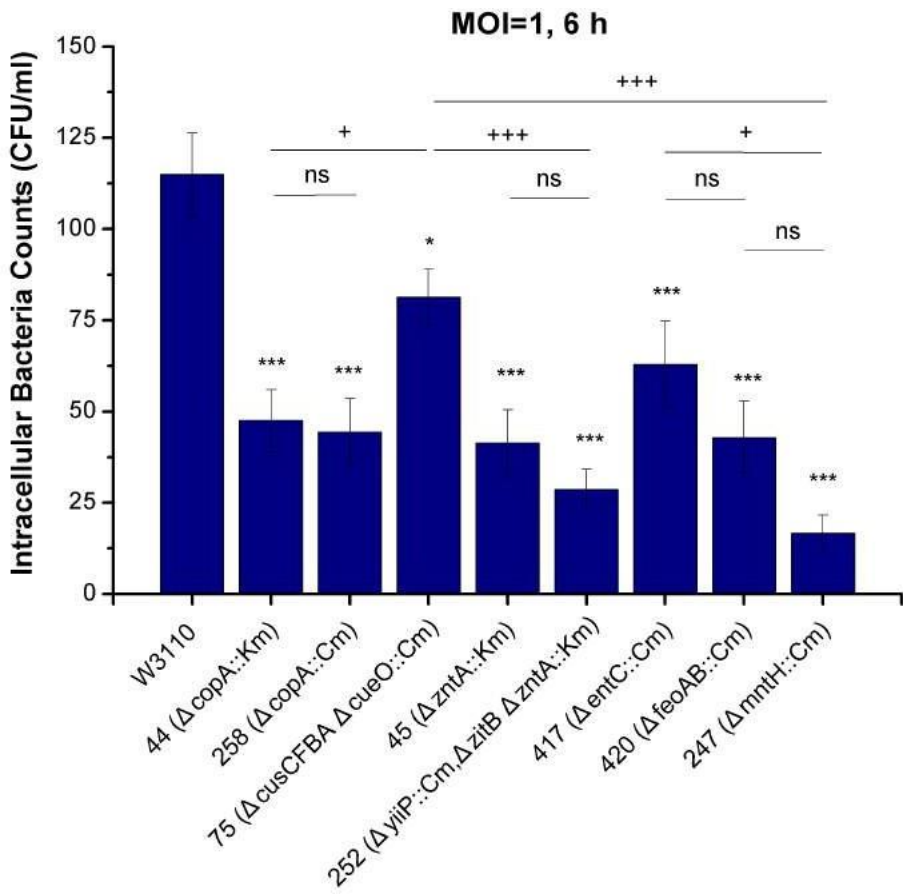
CusF

CusC

P

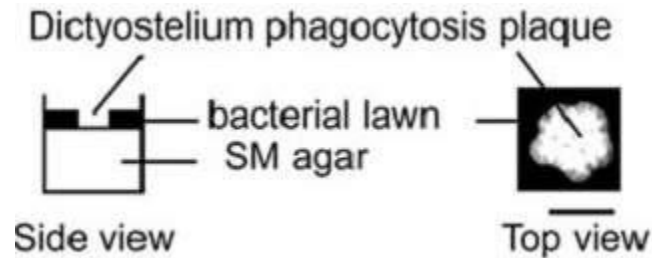
# Cu/Zn exporters and Fe/Mn uptake systems are essential for bacterial survival in *Dictyostelium*

*E. coli*



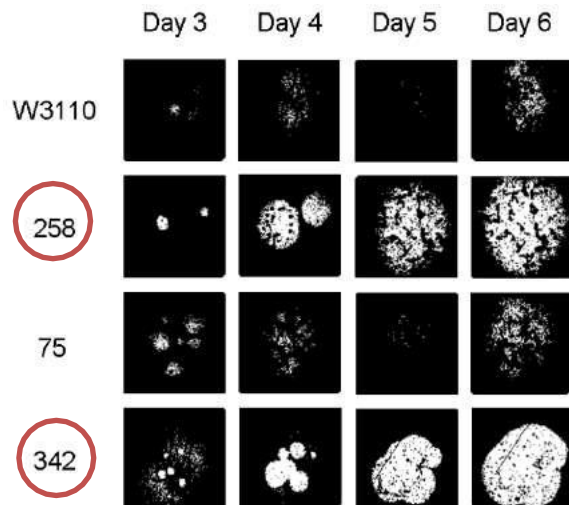
*Δ cueA*

# Survival with different copper resistance determinants towards *Dictyostelium*



Alibaud *et al.* Cellular Microbiology,

***E. coli*, 2.5% HL5-agar**

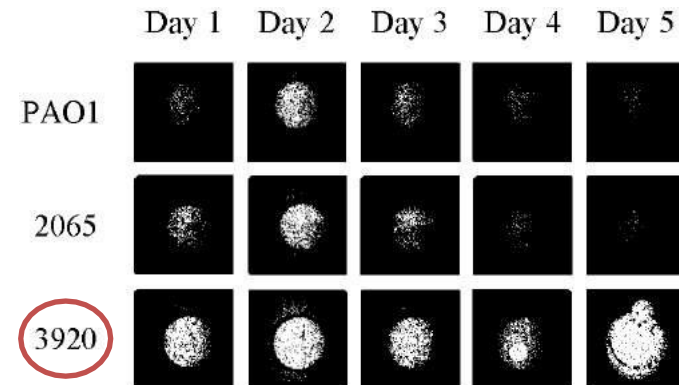


***E. coli* 258:  $\Delta copA::Cm$**

*E. coli* 75:  $\Delta cusCFBA, \Delta cueO::Cm$

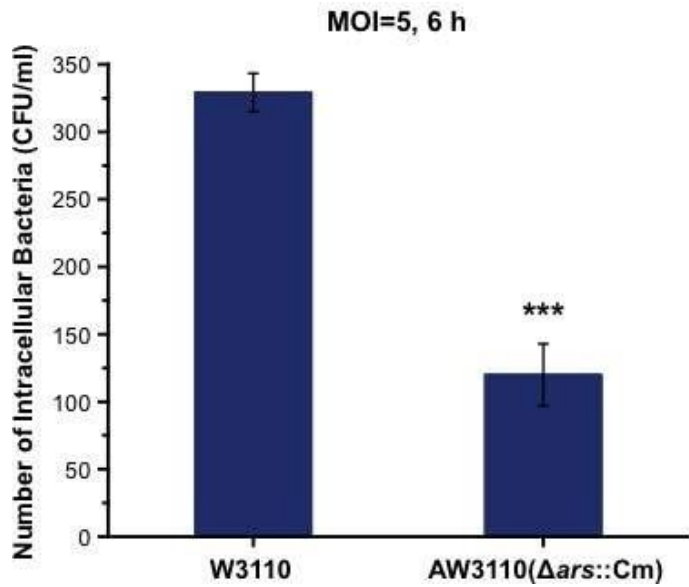
***E. coli* 342:  $\Delta copA::Km, \Delta cusCFBA, \Delta cueO::Cm$**

***Pseudomonas*, 5% HL5-agar**



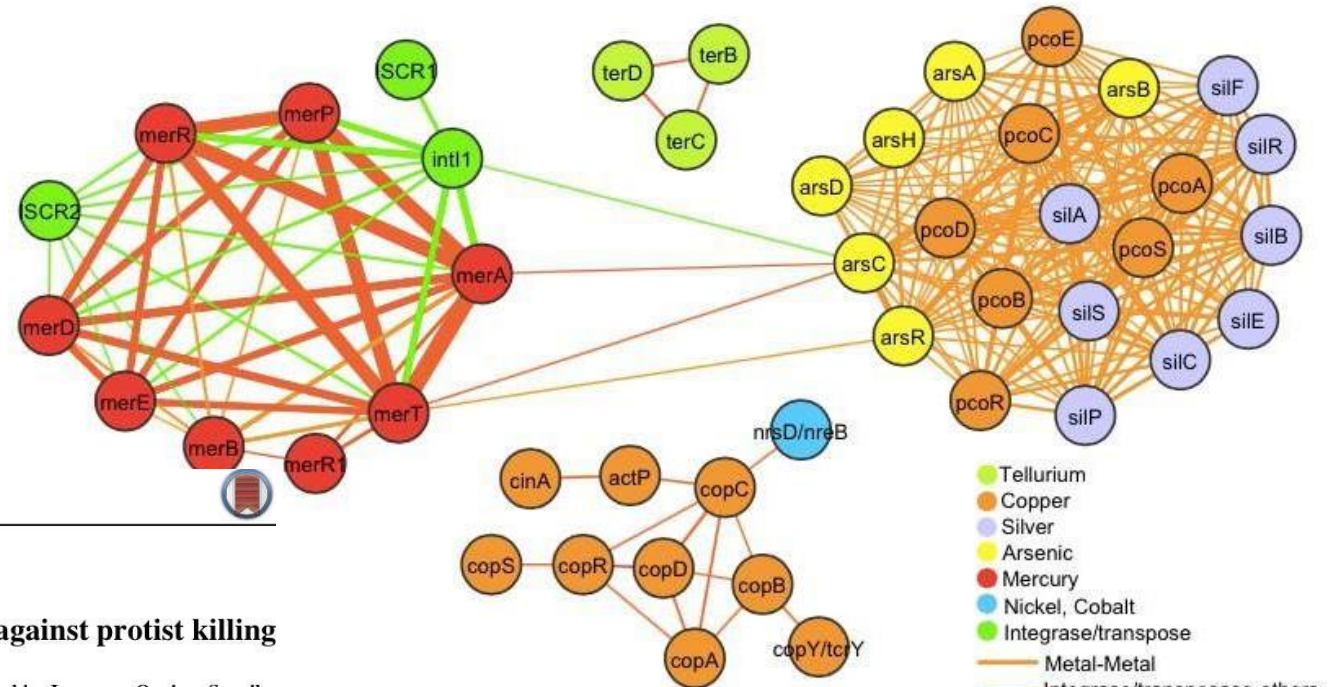
*P. aeruginosa* 2065:  $\Delta pcoA::Cm$

***P. aeruginosa* 3920:  $\Delta cueA$**



Ars resistance often co-occurs on plasmids with copper resistance

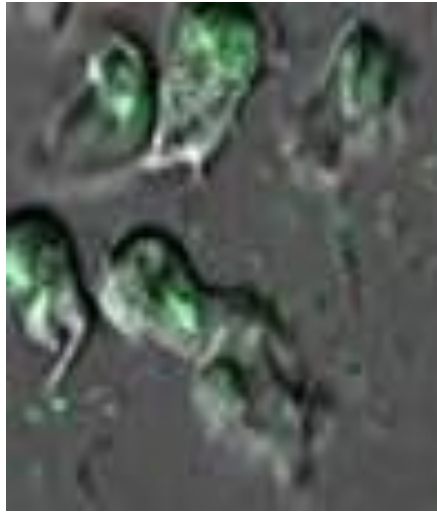
As is used as toxin to kill bacteria



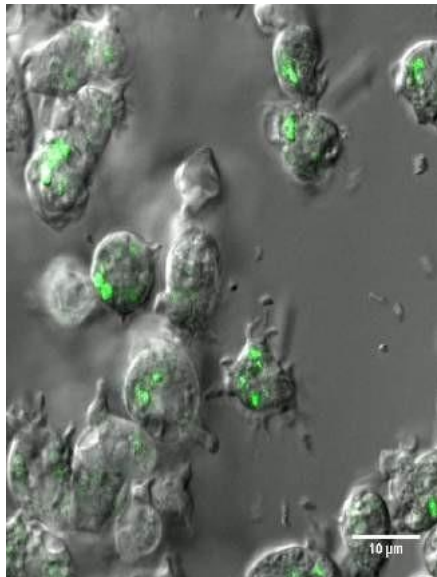
### Bacterial resistance to arsenic protects against protist killing



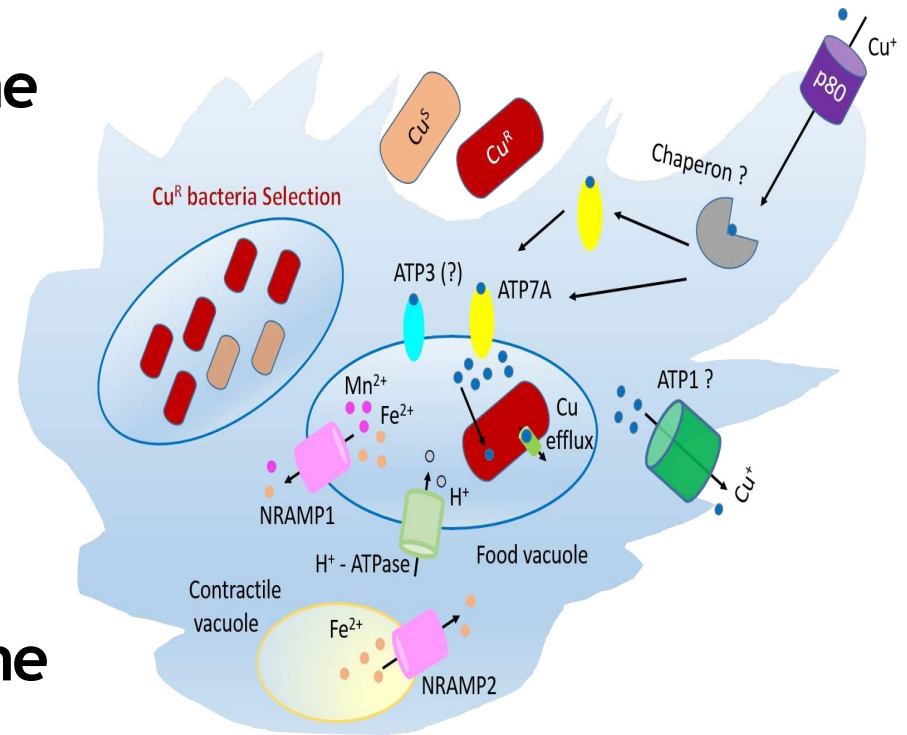
# Zn and Cu dependent expression of gfp



Zn in phagosome



Cu in phagosome



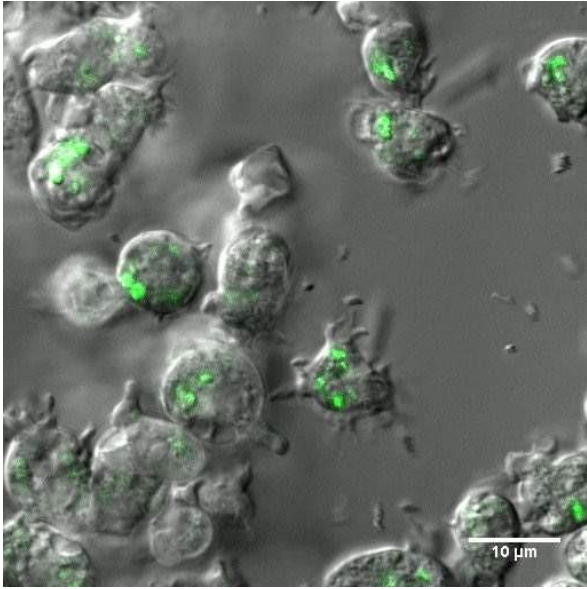
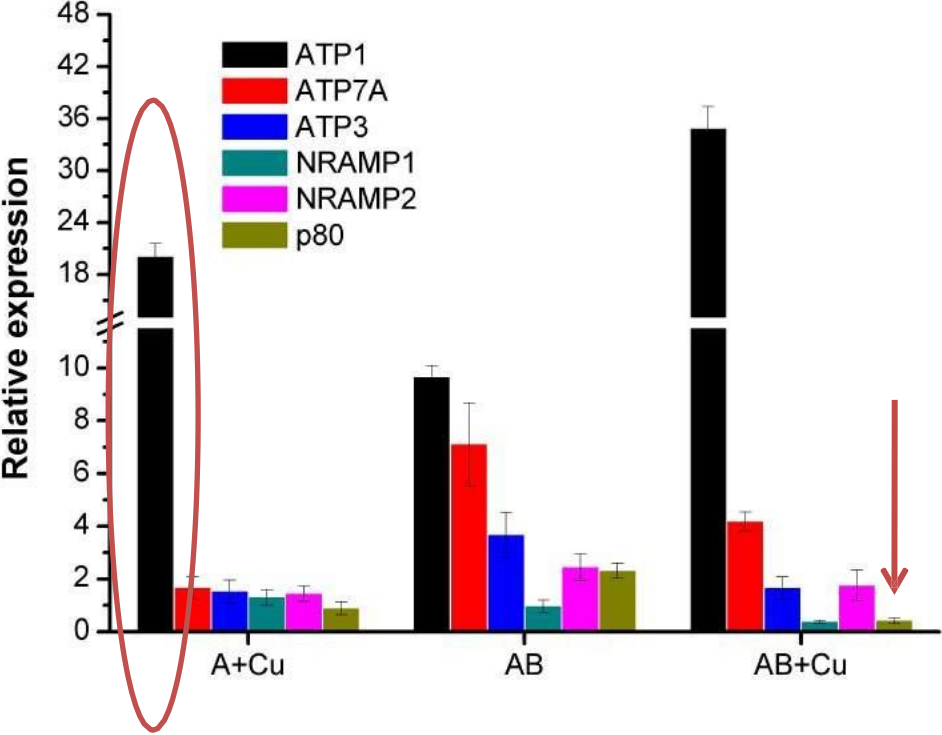
Molecular Microbiology [2016] 102(4), 628–641 ■

doi:10.1111/mmi.13483  
First published online 31 August 2016

**A role for copper in protozoan grazing – two billion years selecting for bacterial copper resistance**

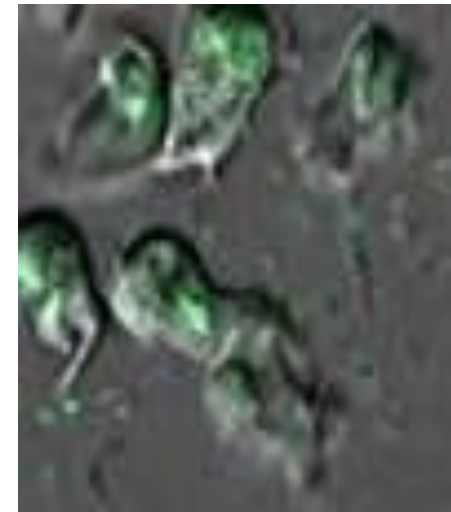
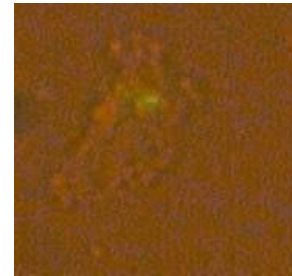
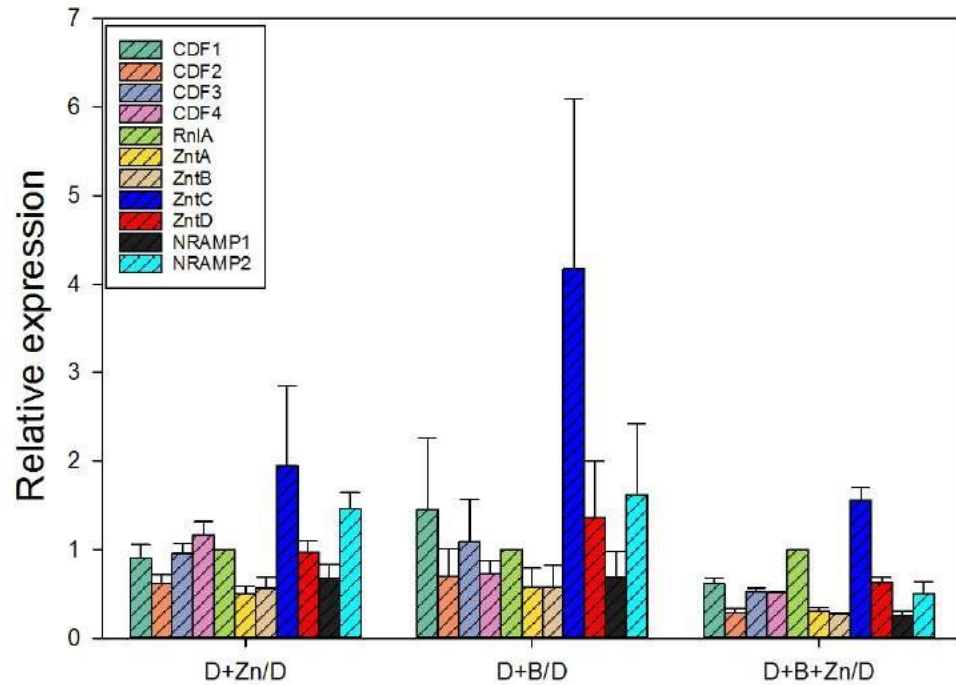


# Cu(I) trafficking was triggered in *Dictyostelium* by bacteria ingestion



*Dictyostelium* + *P.putida* (pCuS-GFP)

# Zn(II) trafficking was triggered in *Dictyostelium* by bacteria ingestion



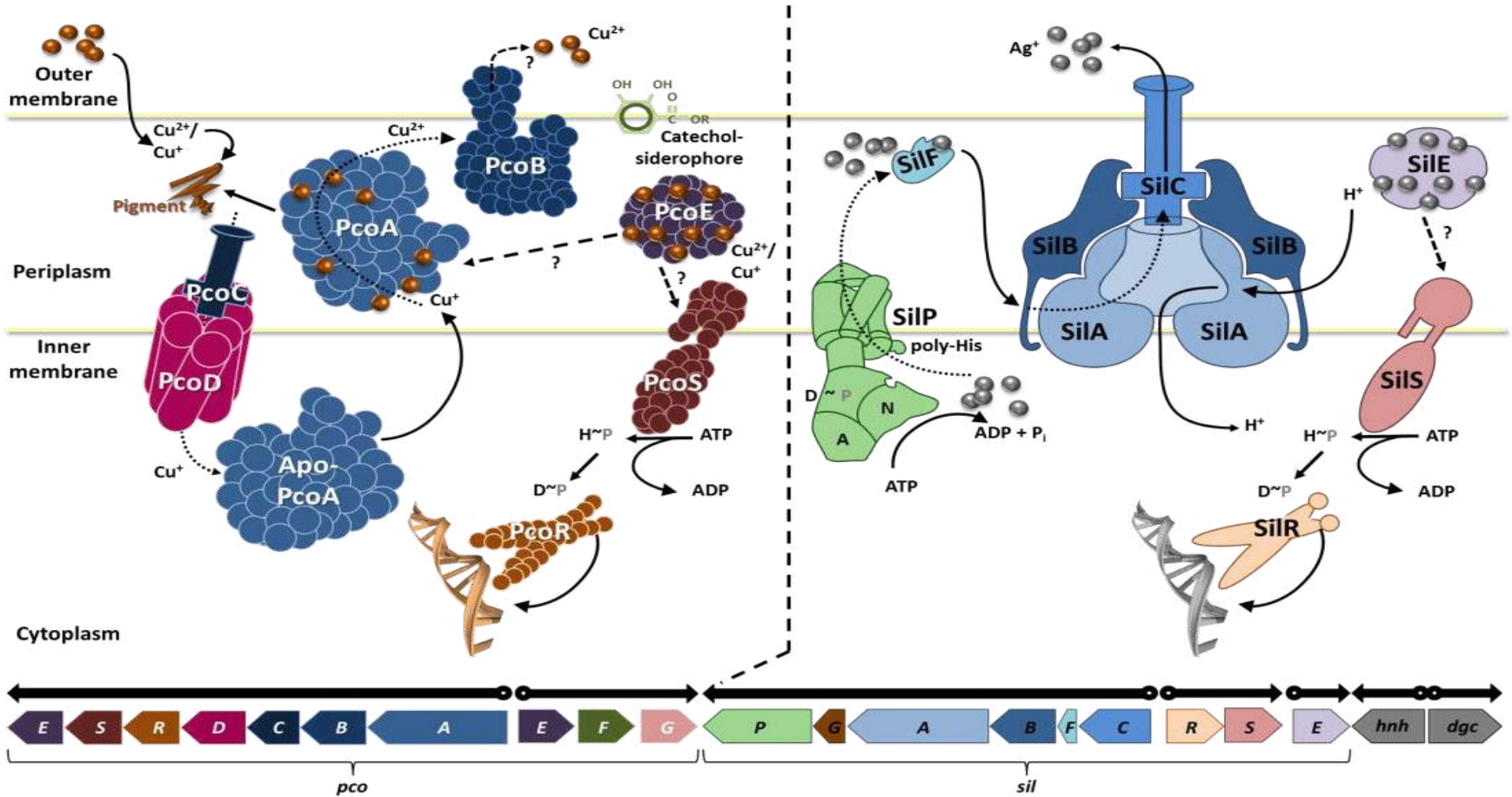
*Dictyostelium* + *M. metallidurans*  
(pCadA-GFP)

# Cu and Zn fed to livestock



Daily feed (kg/day)	Copper (mg/kg diet)	Zinc (mg/kg diet)
Cattle (5.5–10)	10 (8)	30 (8)
Swine (1–2)	100–250 (11)	2000–3000 (11)
Poultry (0.1–0.2)	4–8 (12)	30–60 (12)

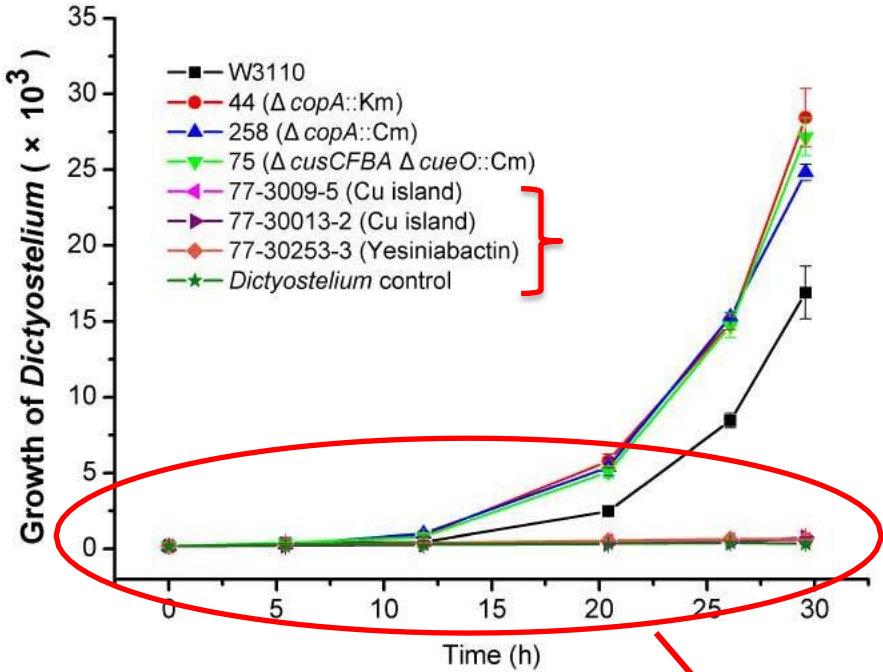
# Cu-resistance & Cu-pathogenicity island?



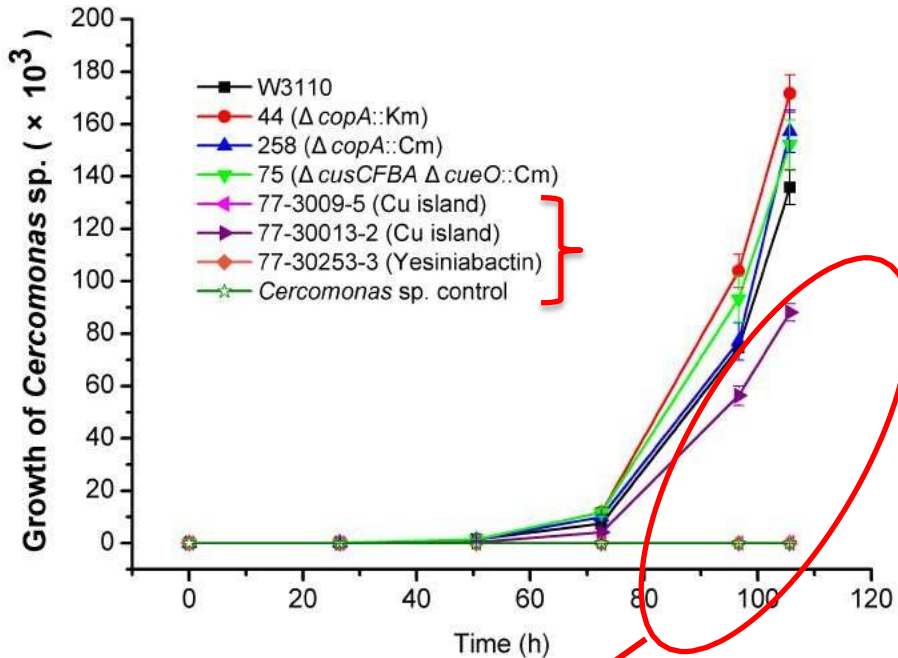
PcoESRDCBAE XX SilP CopG SilABFCRSEX

# Bacterial Cu<sup>R</sup> determinants influence the growth of protozoa

## Dictyostelium



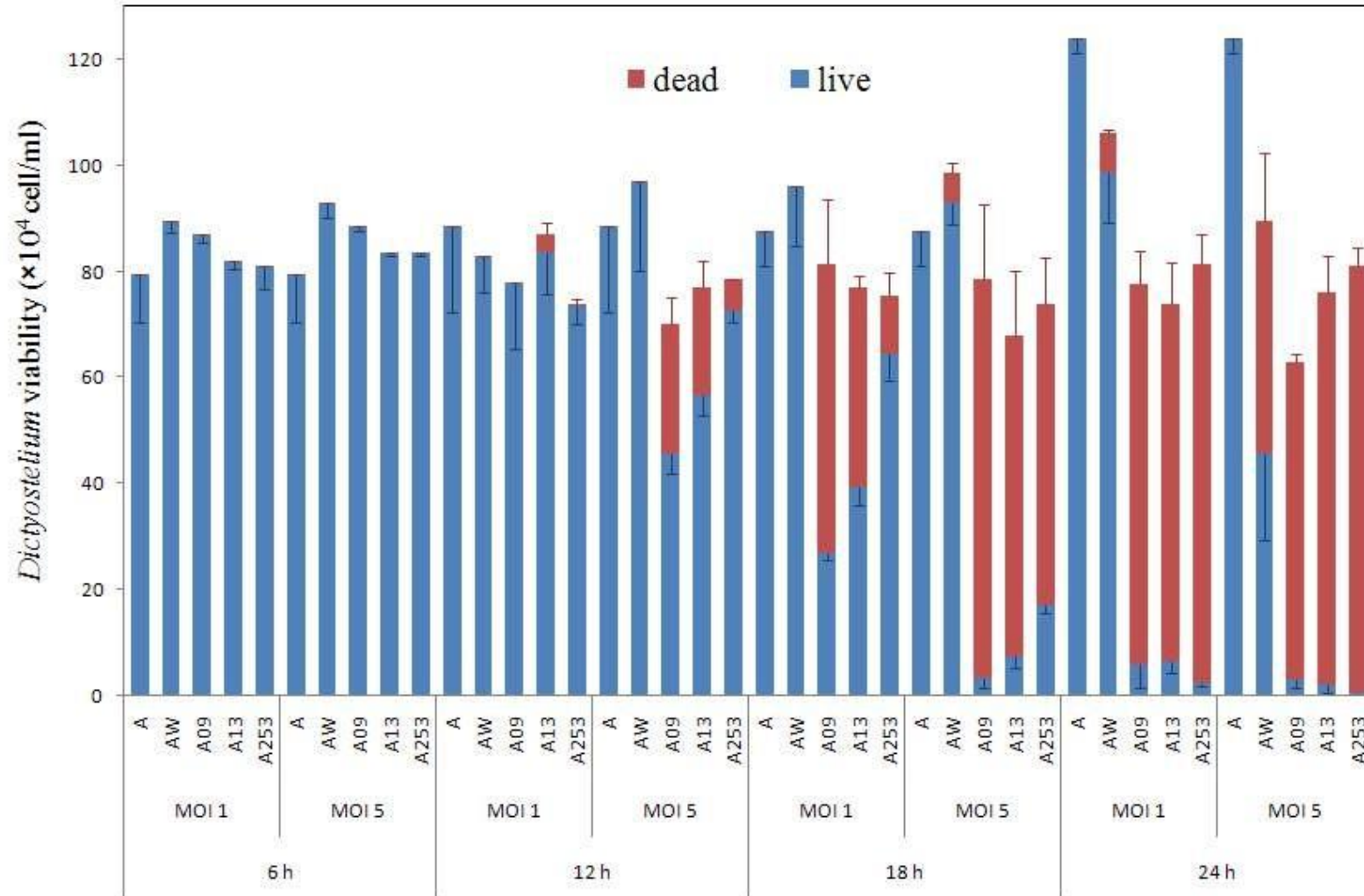
## Flagellate



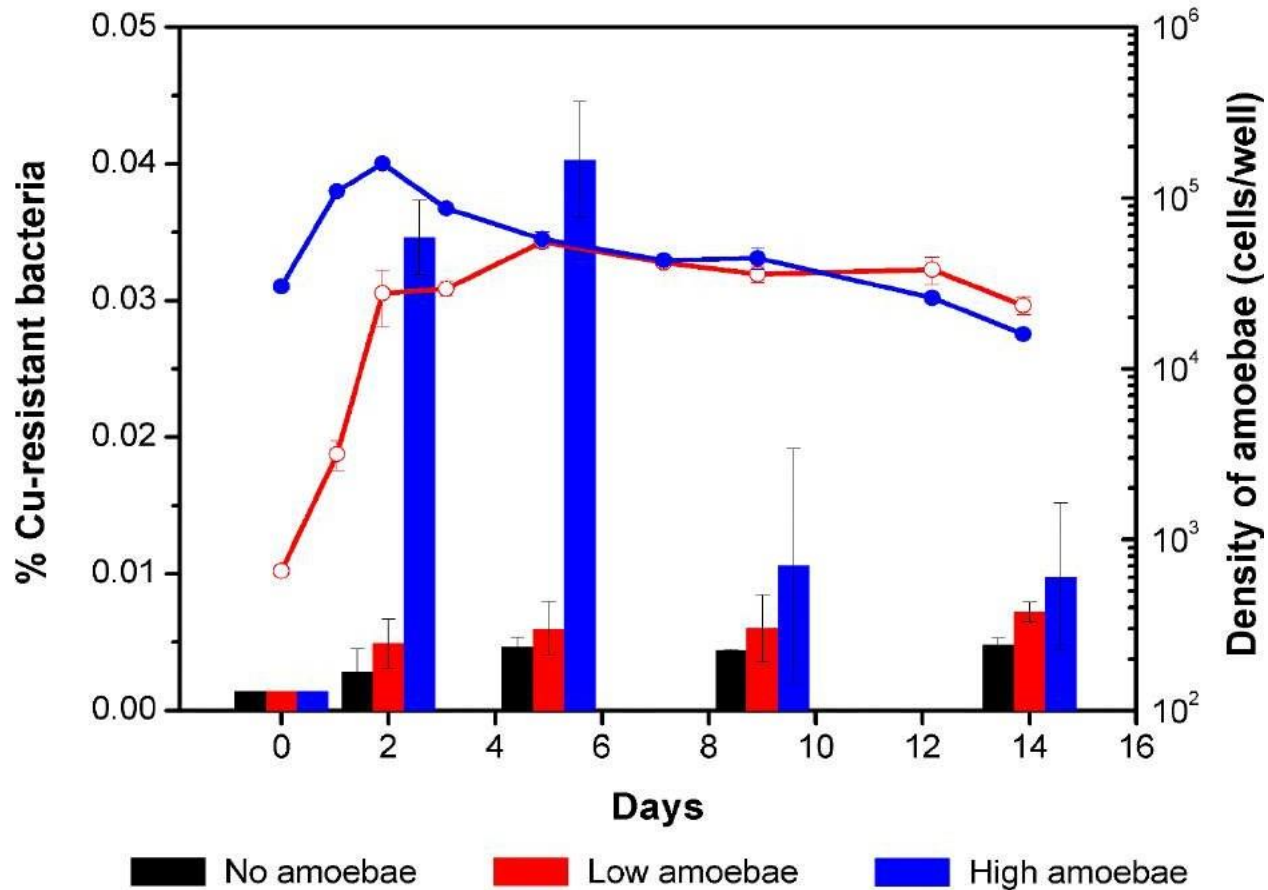
Little growth when fed with Cu<sup>R</sup> strain containing Cu<sup>R</sup> island

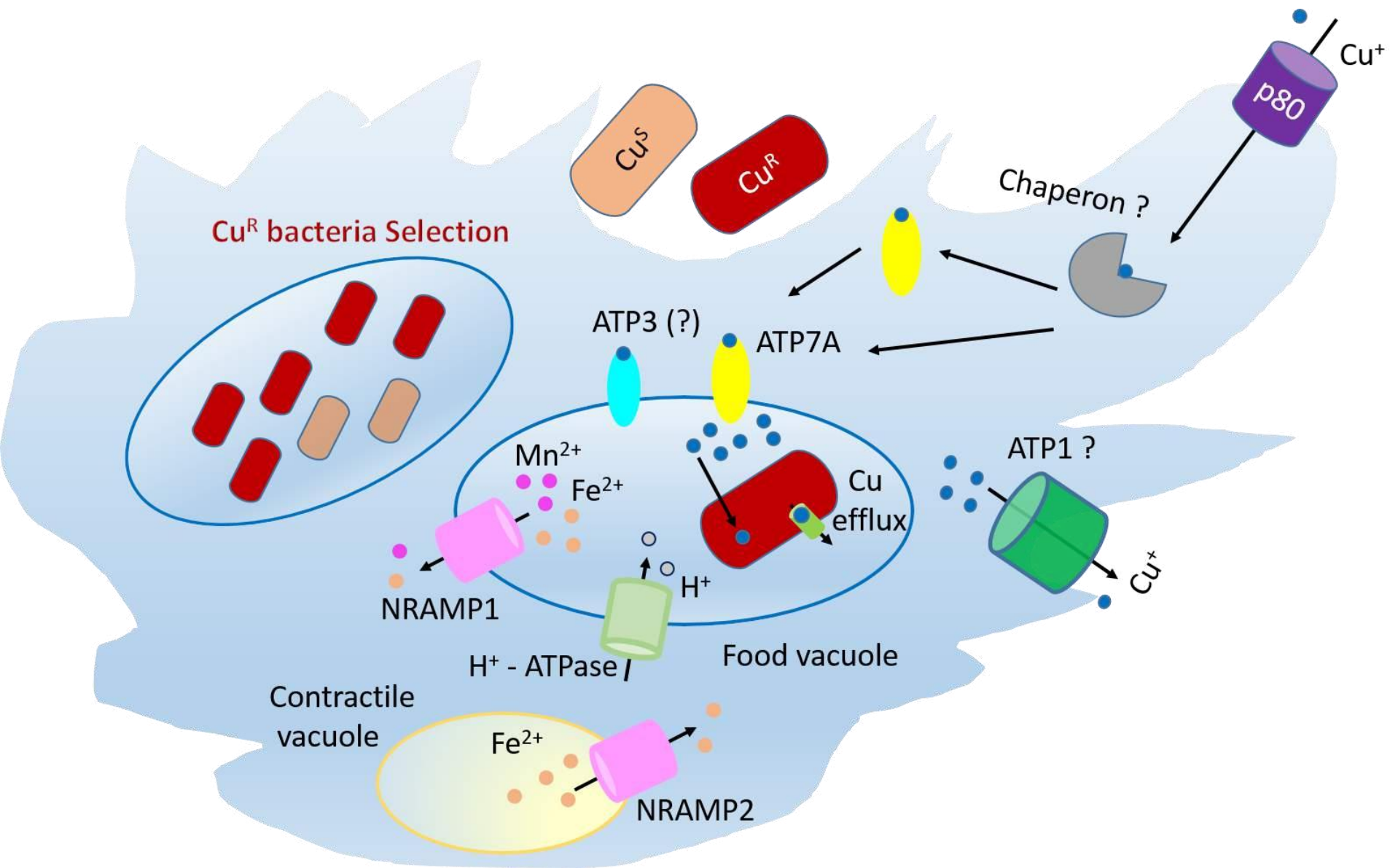


# Dictyostelium viability with W31 10 and three Cu<sup>R</sup> E. coli



# Selection of *Dictyostelium* grazing on Cu-resistant bacteria





## The *cop* operon is required for copper homeostasis and contributes to virulence in *Streptococcus pneumoniae*

Sulman Shafeeq,<sup>3†</sup> Hasan Yesilkaya,<sup>2†</sup>  
Tomas G. Kloosterman,<sup>3</sup> Geetha Narayanan,<sup>1</sup>  
Michal Wandel,<sup>3</sup> Peter W. Andrew,<sup>2</sup> Oscar P. Kulpers<sup>3</sup>  
and Julie A. Morrissey<sup>1\*</sup>

stasis also appears to be required for survival in the nasopharynx.

### Introduction

## Copper resistance is essential for virulence of *Mycobacterium tuberculosis*

Frank Wolschendorf<sup>a</sup>, David Ackart<sup>b</sup>, Tej B. Shrestha<sup>c</sup>, Laurel Hascall-Dove<sup>b</sup>, Scott Nolan<sup>d</sup>, Gyanu Lamichhane<sup>d</sup>, Ying Wang<sup>a</sup>, Stefan H. Bossmann<sup>c</sup>, Randall J. Basaraba<sup>b</sup>, and Michael Niederweis<sup>a,1</sup>

<sup>a</sup>Department of Microbiology, University of Alabama at Birmingham, Birmingham, AL 35294; <sup>b</sup>Department of Microbiology, Immunology, and Pathology, Colorado State University, Fort Collins, CO 80523-1619; <sup>c</sup>Department of Chemistry, Kansas State University, Manhattan, KS 66506-0401; and <sup>d</sup>Tuberculosis Animal Research and Gene Evaluation Taskforce, The Johns Hopkins University School of Medicine, Baltimore, MD 21231

Edited by Emil C. Gotschlich, The Rockefeller University, New York, NY, and approved December 6, 2010 (received for review June 30, 2010)

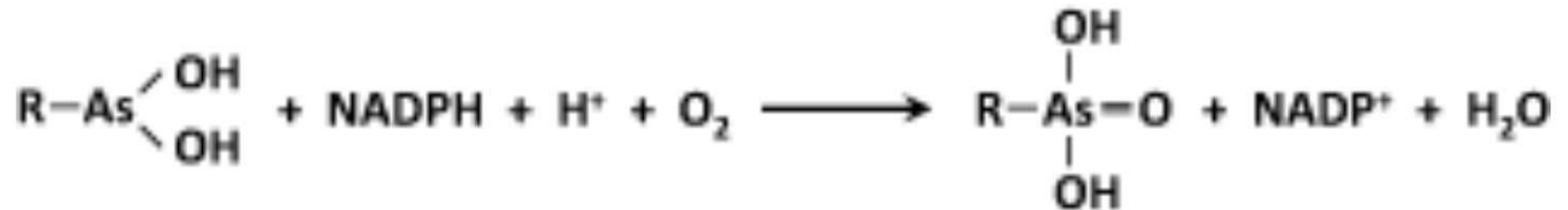
## Role of Copper Efflux in Pneumococcal Pathogenesis and Resistance to Macrophage-Mediated Immune Clearance

Michael D. L. Johnson,<sup>a</sup> Thomas E. Kehl-Fie,<sup>b</sup> Roger Klein,<sup>a\*</sup> Jacqueline Kelly,<sup>a\*</sup> Corinna Burnham,<sup>a</sup> Beth Mann,<sup>a</sup> Jason W. Rosch<sup>a</sup>

Department of Infectious Diseases, St. Jude Children's Research Hospital, Memphis, Tennessee, USA<sup>a</sup>; University of Illinois Urbana-Champaign, Department of Microbiology, Urbana, Illinois, USA<sup>b</sup>

## Warum *merA*, *arsH*, *ter* operon?

NADPH-dependent FMN reductase ArsH



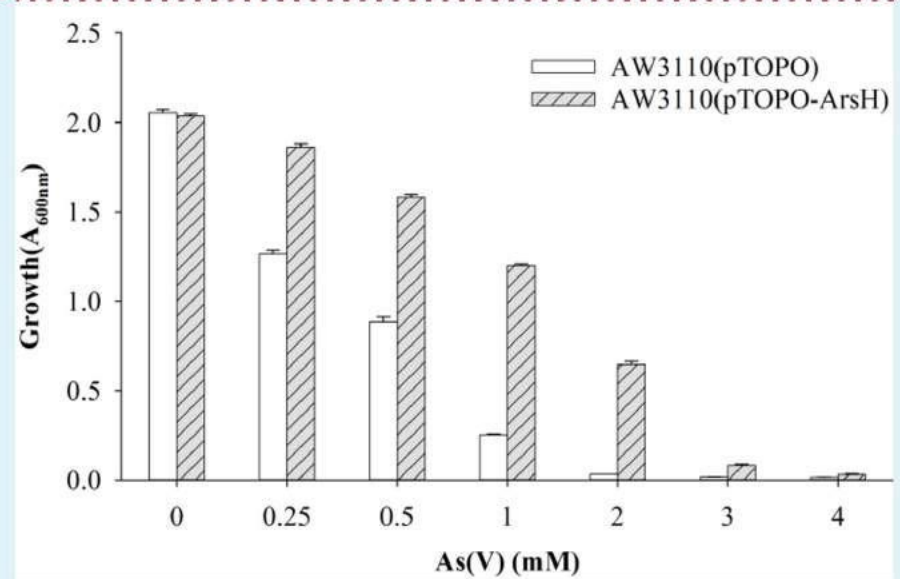
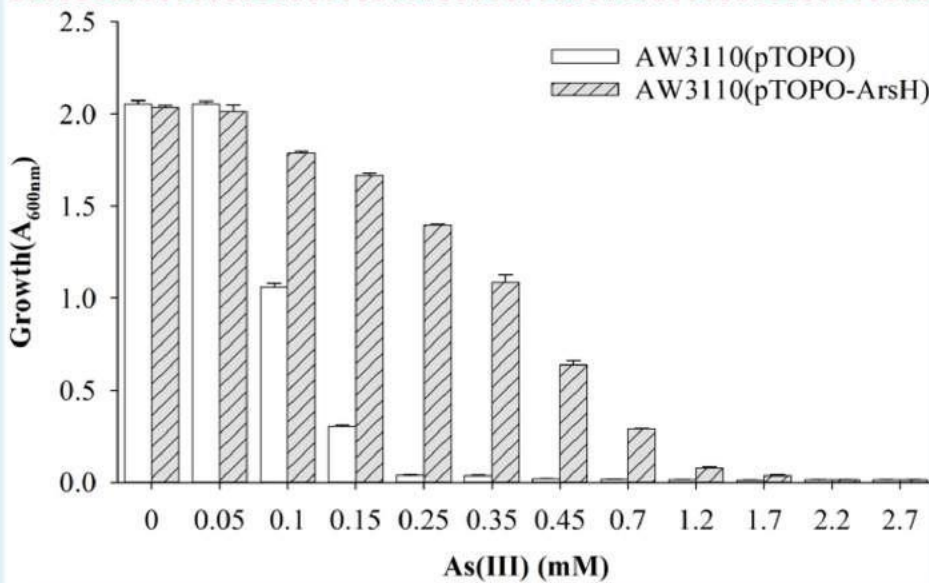
MMA(III) ist wesentlich toxischer als As(III)



# Expression von ArsH macht Zellen resistenter gegen As(III), As(V), Sb(III) und Sb(V)

**(A)**

**(B)**



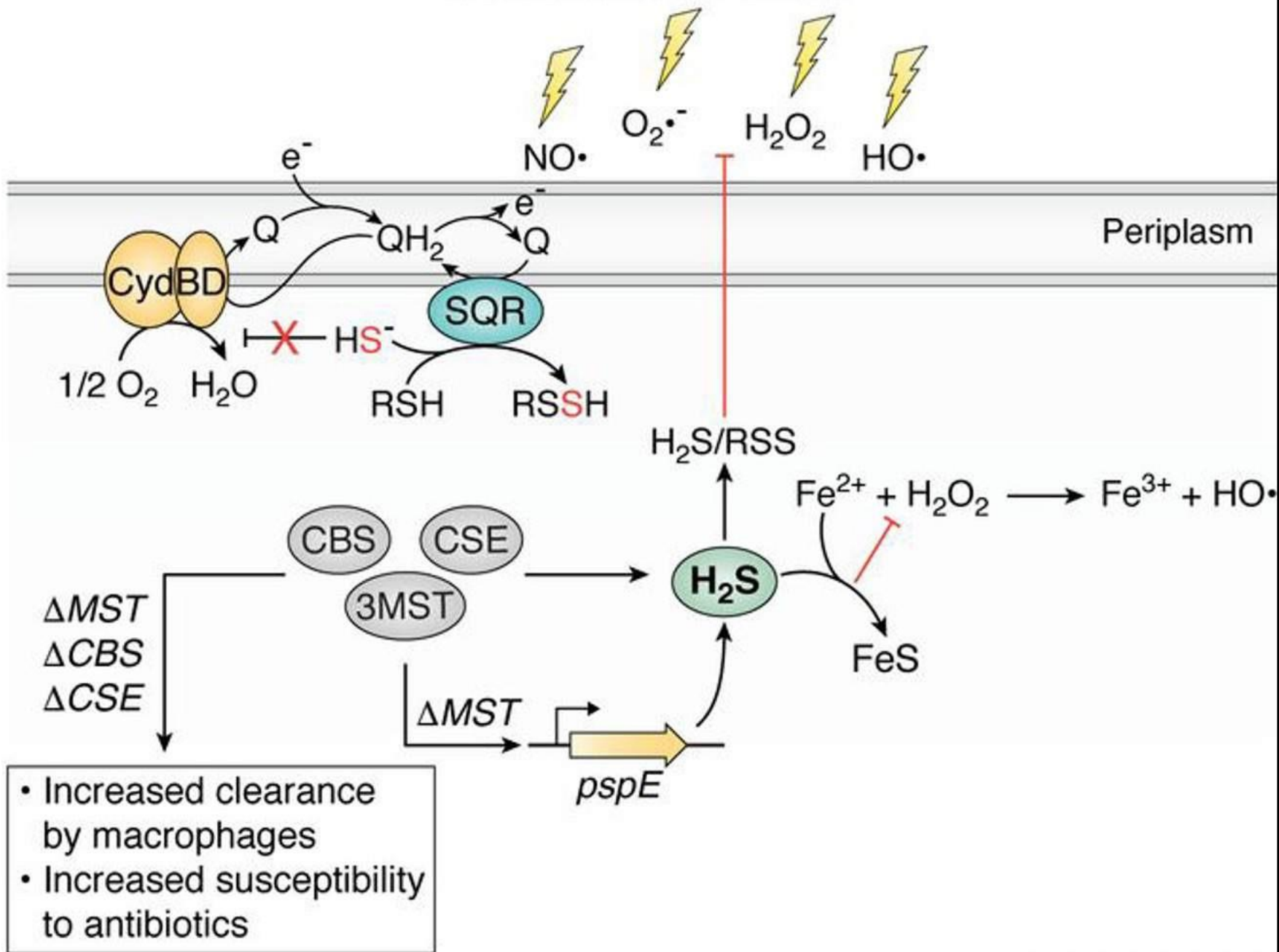
# **ArSH protects *Pseudomonas putida* from oxidative damage caused by exposure to arsenic**

*ArSH1 and ArSH2 protect P. putida from oxidative stress caused by diamide*

Diamide thiol-specific oxidizing agent, which reacts with thiols and protein sulfhydryls, thereby oxidizing glutathione and promoting formation of disulfide bonds

**B**

## Innate immune response



# *Staphylococcus aureus* responds to allicin by global S-thioallylation – Role of the Brx/BSH/YpdA pathway and the disulfide reductase MerA to overcome allicin stress

Vu Van Loi<sup>a</sup>, Nguyen Thi Thu Huyen<sup>a,1</sup>, Tobias Busche<sup>a,b</sup>, Quach Ngoc Tung<sup>a</sup>, Martin Clemens Horst Gruhlke<sup>c</sup>, Jörn Kalinowski<sup>b</sup>, Jörg Bernhardt<sup>a,d</sup>, Alan John Slusarenko<sup>c</sup>, Haike Antelmann<sup>a,\*</sup>

<sup>a</sup> Leibniz Universität Hannover, Institute of Microbiology, 30559 Hannover, Germany

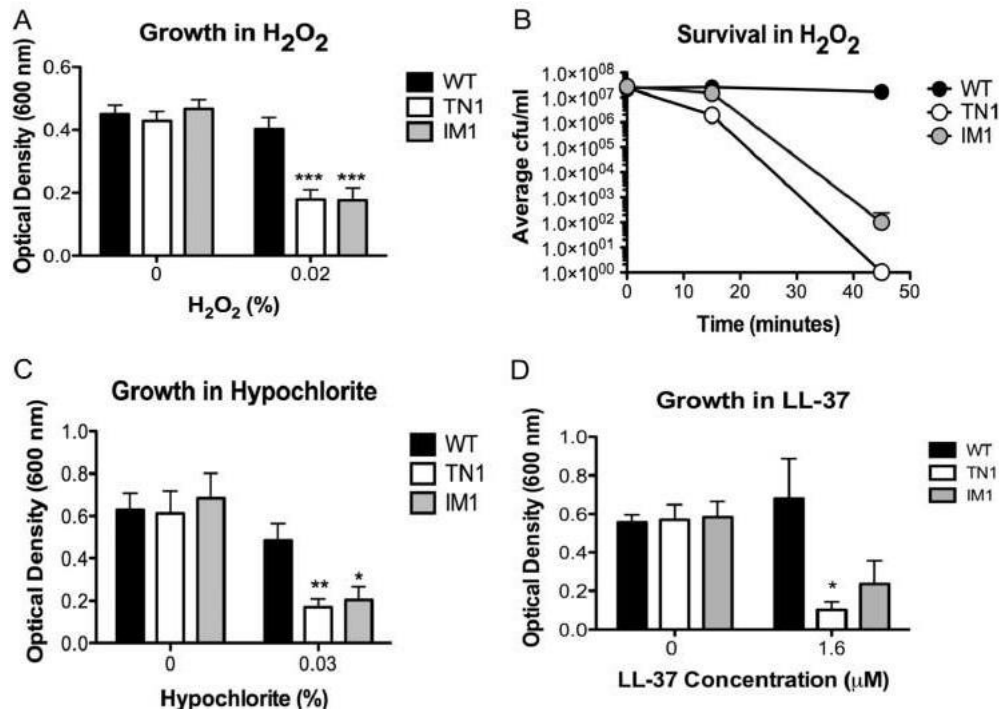
Allicin provokes a strong thiol-specific oxidative and sulfur stress response in the *S. aureus* USA300 transcriptome

Biochemical assays indicate that MerA functions in allicin detoxification, the HypR-controlled disulfide reductase MerA

# Novel Role for the *yceGH* Tellurite Resistance Genes in the Pathogenesis of *Bacillus anthracis*

Sarah E. Franks,<sup>a</sup> Celia Ebrahimi,<sup>b</sup> Andrew Hollands,<sup>b</sup> Cheryl Y. Okumura,<sup>b</sup> Raffi V. Aroian,<sup>c</sup> Victor Nizet,<sup>b,d</sup> Shauna M. McGillivray<sup>a</sup>

Department of Biology, Texas Christian University, Fort Worth, Texas, USA<sup>a</sup>; Department of Pediatrics,<sup>b</sup> Division of Biological Sciences,<sup>c</sup> and Skaggs School of Pharmacy and Pharmaceutical Sciences,<sup>d</sup> University of California San Diego, La Jolla, California, USA





Metall Resistenzen schuetzen vor Vergiftung durch Cu(I), Zn(II) und As(III), sind Virulenzfaktoren

Nebenjob der Metall Resistenzen ist der Schutz vor reaktiven Sauerstoffverbindungen

Selektionsvorteil auch ohne Vorhandensein von Metallen, Selektionsvorteil